Antioxidant and cytotoxic properties of *Pavetta crassicaulis* Bremek. leaf crude extract and its isolated pure compound

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Pavetta crassicaulis Bremek. (Family Rubiaceae), an important ethnomedicinal shrub/small tree, belonging to the family Rubiaceae, grows up to 4 meters tall. The plant is endemic to peninsular India. Plant parts have many medicinal properties used in the treatment of arthritis, boils and itches, hemorrhoidal pains, dropsy, epilepsy, urinary complaints, and anticephalagic. Fruits used as anthelmintic and flowers are eaten fried. The antioxidant experiments conducted were metal chelating activity, superoxide radicals, hydroxyl radical, DPPH radicals, ABTS radical scavenging assays. *In vitro* cytotoxicity was tested in DLA and EAC cancer cells by using Trypan blue dye exclusion technique and MTT assay. The results revealed that the leaf ethanolic crude extract of the *Pavetta crassicaulis* Bremek. has appreciable radical scavenging activity and its extracted pure compound, 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol, showed appreciable radical scavenging activity but not as good as the standards used. The cytotoxic experiment revealed that leaf ethanolic extract has considerable cytotoxicity and its pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol also showed noticeable activity which is not comparable to the standard Curcumin.

Keywords: Antioxidant, Cytotoxic, DLA, *Pavetta crassicaulis* Bremek, EAC, 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol.

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Introduction

Most of the free radicals divided into derivatives of oxygen like superoxide, hydrogen peroxide and derivatives of nitrogen like nitric oxide and peroxynitrite were an influence on biological systems¹.

Reactive oxygen species induce toxic effects like inactivation of enzymes and alteration of intracellular oxidation-reduction state². It can influence the DNA modifications and chromosome aberrations leading to carcinogenesis^{3,4}.

The free radicals damage on the cell/tissues is neutralised by antioxidants such as á-tocopherol, carotenoids, glutathione, thiols, vitamin C etc., by scavenging and decreasing their formation. Most of the plants have several natural compounds exhibiting antioxidant and/or radical scavenger properties⁵. There is clear evident proof that most of the phenolic compounds present in the plant possess antioxidant properties⁶.

Many of the plant extracts are extensively used as medicinal compounds for the treatment of various ailments in different parts of the world, especially in under developing and developing countries⁷⁻⁹.

The Indian subcontinent is a vast repository of medicinal plants that are used in traditional medical treatments which also forms a rich source of knowledge¹⁰. India is one of the megadiversity centres on the planet having a diverse medicinal plant species, which is unexplored and most of them are endemic. India shares approximately 13 % of world's biodiversity, one among 17 mega diversity centres. Among the 34 hotspots in the world, India has four hotspots, viz., Eastern Himalaya, Indo-Burma, Western Ghats, and the Andaman and Nicobar Islands. The various indigenous systems use several plant species to treat different ailments¹¹. In India, around 20,000 medicinal plant species have been recorded recently, but more than 500 traditional communities use about 800 plant species for curing different diseases¹².

Experimental medicinal plant and description

Pavetta crassicaulis Bremek. is an important ethnomedicinal shrub or small tree belonging to the family Rubiaceae. It grows up to four meters tall and is endemic to peninsular India. The leaves are often membranous with dark bacterial nodules, having small white tubular flowers, sometimes salviform or funnelshaped with four spreading petal lobes. The flowers are carried on terminal corymbs or cymes, the leaves are elliptical-oblong to elliptic-lanceolate, 6-15 cm long, and pointed at both ends. The flowers are white, rather fragrant, and borne in considerable number in hairy terminal panicle which is 6-10 cm long. The sepals are very small and toothed. The flowers tube is slender and about 1.5 cm long, with obtuse petals above half the length of the tube. The fruits are black when they dry, somewhat rounded and about 6 mm in diameter¹³.

P. crassicaulis Bremek. is endemic to peninsular India and distributed in many states like, Mizoram, Gujarat, Uttara Pradesh, Chhattisgarh, Orissa Maharashtra and Karnataka¹⁴⁻¹⁶.

The plant has medicinal properties used in the treatment of arthritis, boils and itches, hemorrhoidal pains, visceral problems, dropsy¹⁷, epilepsy, hemorrhoids¹⁸, skin diseases¹⁹, anticephalagic, fat burner, aphrodisiac²⁰, urinary complaints and fruits used as anthelmintic and flowers are eaten fried²¹.

The preliminary qualitative phytochemical analysis of *P. crassicaulis* Bremek. methanolic crude extract revealed the presence of alkaloids, steroids and terpenoid with moderate antimicrobial activity²²⁻²³.

Despite many works on *P. crassicaulis* Bremek, antioxidant and cytotoxicity properties have not been studied in detail. Therefore, the study aimed to provide basic data on the antioxidant and cytotoxic study of *P. crassicaulis* Bremek plant parts.

Materials and Methods

Plant collection and authentication

The leaf of *P. crassicaulis* Bremek. was collected from Shringeri taluk, Karnataka in April 2014.(13.4198° N, 75.2567° E) The plant was identified by Prof. K G Bhat, Udupi and a voucher specimen was conserved under the reference number KU/AB/RN/AS/002.

Extraction

The samples were dried in the shade for 20 to 25 days, mechanically powdered and subjected to Soxhlet extraction using ethanol and aqueous extracts²⁴. The crude extracts were collected in airtight plastic containers and stored in cool condition.

GC-MS analysis

Plant extracts were subjected to gas chromatography-mass spectrometry (GC-MS); obtained spectra were analysed. GC Model: Thermo Trace GC Ultra, MS Model: Thermo DSQ II, Ionization: Electron Impact Ionisation (EI), Chemical Ionisation (CI), Mass Range: 1 - 1074 m/z.

Chemicals used

2,2-Diphenyl-2-picrylhydrazyl(DPPH), 2,2-azinobis(3-ethylbenzothiazoline)-6-sulfonic acid (ABTS), ascorbic acid, butylated hydroxyl anisole (BHA), Ferrozine, gallic acid, ferrous chloride, Folin– Ciocalteu reagent, nitro blue tetrazolium sodium salt (NBT), Nicotinamide adenine dinucleotide phosphate reduced (NADH), phosphate buffered saline (PBS) and trichloroacetic acid (TCA), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide, Trypan blue. All other chemicals and solvents used were of analytical grade.

Qualitative phytochemical screening for phenols and flavonoids *Total phenolic content*

Total phenolic content was determined by the Folin-Ciocalteau method²⁵. A dilute concentration of extract (0.5 mL) was mixed with 0.5 mL of 1:1 diluted Folin-Ciocalteu reagent and 4 mL of sodium carbonate (I M). The mixtures were allowed to stand for 15 min, and the absorbance was measured against the blank at 765 nm colourimetrically. A standard curve was plotted using different concentrations of Gallic acid (standard, 0-1000 μ g/mL of ethanol). The total phenolic content of solvent extracts was estimated as μ g Gallic acid equivalents (GAE)/mg of extract. All samples were analysed in triplicate.

Determination of total flavonoid content

Total flavonoid content was determined²⁶ using quercetin as standard. Briefly, 5 mL of extracts (200 μ g) in Millipore water was mixed with 300 μ L of sodium nitrite (5 %) and 300 μ L of aluminum chloride (10 %), this reaction mixture was incubated at room temperature for 6 min followed by the addition of 2 mL of sodium hydroxide (1 M). Later, the volume in each test tube was made up to 10 mL by adding 2.4 mL of Millipore water. Absorbance was measured against the blank at 510 nm. Total flavonoid content of the extract was expressed in terms of equivalent to Quercetin (EQ, μ g/mg of dry mass).

In vitro antioxidant activity

Total antioxidant capacity

Total antioxidant capacity²⁷ of extracts were analysed using 300 μ L of extracts at different concentrations (0-160 μ g) was combined with 3 mL of reagent mixture (4 mM ammonium molybdate, 0.6 M sulfuric acid and 28 mM of sodium phosphate). The tubes were capped and kept for incubation at 95 °C for 90 min, after cooling to room temperature, the absorbance of the content was measured at 695 nm against the blank. The total antioxidant capacity of each extract is expressed as equivalents of ascorbic acid.

Total reductive capability

Total reduction capability²⁸ of extracts was analysed using different concentration of extracts (0-300 μ g) in 1 mL of water was mixed with 2.5 mL of phosphate buffer (0.2 M, pH 6.5) and 2.5 mL of potassium ferricyanide (1 %). The mixtures were incubated at 50 °C for 20 min, and 2.5 mL of 10 % trichloro acetic acid were added. Then the mixture was centrifuged for 10 min at 3000 rpm. 2.5 mL of water and 0.5 mL of ferric chloride (0.1 %) was added to 2.5 mL of supernatant. The absorbance was measured at 700 nm against blank. A higher absorbance of the reaction mixture indicated greater reducing power. Total reducing capacity of each extract is expressed as equivalents of quercetin.

Scavenging of superoxide radicals

Superoxide radical scavenging activity was determined by the NBT reduction method²⁹. The reaction mixture contained 6 μ M EDTA, 0.0015 % NaCN, 2 μ M riboflavin, 50 μ M NBT, various concentrations of extract, and phosphate buffer (67 mM, pH 7.8) in a final volume of 3 mL. The tubes were uniformly illuminated with an incandescent lamp for 15 min, and the optical density was measured at 560 nm before and after illumination. The percentage inhibition of superoxide radical generation was evaluated by comparing the absorbance values of control and experimental tubes.

Scavenging of hydroxyl radical

Hydroxyl radicals generated from Fe²⁺/ascorbate/H system degrades deoxyribose producing thiobarbituric acid reacting substance (TBARS)³⁰. The efficacy of the extracts to inhibit TBARS formation was assessed. The reaction mixture contained 2.8 mM deoxyribose, 0.1 mM FeCl₃, 0.1 mM EDTA, 1 mM H₂O₂, 0.1 mM ascorbic acid, 20 mM KH₂PO₄–KOH (pH 7.4), and various concentrations of extracts in a final volume of 1 mL. The reaction mixture was incubated for 1 h at 37 °C. The TBARS formed was measured³¹ and the percentage inhibition was calculated from the optical measurements of control and experimental tubes.

Scavenging of DPPH radicals

Stable radical, 2, 2-diphenyl-1-picryl hydrazyl (DPPH) in methanol was used as a substrate to evaluate antioxidant activity. The method is based on the reduction of DPPH radical in the presence of hydrogen donating antioxidants leading to the formation of a non-radical form DPPH-H by the reaction. DPPH in its radical form has an absorption peak at 515 nm which disappeared upon reduction by antioxidant compounds. Absorbance was measured 20 min after the reaction was started.

Radical scavenging activity was calculated using the following formula

$$Percentage inhibition = \frac{Absorbance of control - Absorbance of test}{Absorbance of control} \times 100$$
... (1)

IC₅₀ value was calculated using the following formula

$$IC_{50} = \frac{Sum of extract concentration}{Sum of percentage of inhibition at diff conc.} \times 50$$
....(2)

Scavenging of ABTS radicals

ABTS (2, 2-azobis-3-ethylbenthiozoline-6-sulfonic acid) radical scavenging activity³² of the extract was determined based on the principle that the oxidation of ABTS to its cation radicals by ferryl myoglobin is formed in the reaction of H_2O_2 and metmyoglobin. Briefly, the stock solutions of 500 μ M ABTS diammonium salt, 400 μ M myoglobin (MbIII), 740 μ M potassium ferricyanide, and 450 μ M H_2O_2 were prepared in PBS (pH 7.4). Metmyoglobin was prepared by mixing equal volumes of myoglobin and potassium ferricyanide solutions. The reaction mixture (2 mL) contained ABTS (150 μ M), MbIII (2.25 μ M), and varying concentrations of extracts in PBS. The reaction was initiated by adding 75 μ M H_2O_2 and oxidation reaction was monitored at 734 nm.

Metal chelating activity

The chelation of ferrous ions³³ experiment was done with 3 mL of extracts at different concentrations taken in different test tubes followed by the addition of 50 μ L of ferrous chloride (2 mM). The reaction was initiated by the addition of 20 μ L ferrozine (5 mM), and then the mixture was shaken vigorously and allowed to stand for 10 min at room temperature. After equilibrium, absorbance of the solution was measured at 562 nm against the blank. EDTA was used as standard for comparison. Percentage of inhibition and IC₅₀ value was calculated using equation (1) and equation (2).

In vitro cytotoxicity assay

Cell lines

EAC (Ehrlich's Ascites Carcinoma): Paul Ehrlich found the initial tumour for the Ehrlich's Ascites carcinoma in 1905. The ascites variant was obtained in 1932 by intraperitoneal transplantation of Ehrlich's solid adenocarcinoma.

DLA (Dalton's Lymphoma Ascites): The initial tumour for the DLA arose as a Spontaneous Carcinoma in the thymus of mice in 1947. The cell lines were obtained from Amala Cancer Research Centre, Thrissur.

Trypan blue dye exclusion technique

Any compound, which is cytotoxic to cells, inhibits the cell proliferation and kills the cells. Trypan blue³⁴ can penetrate into the dead cells and give it a blue colour. This method gives an exact number of dead and viable cells³⁵.Cells were aspirated from the peritoneal cavity of tumor-bearing mice, and it was washed three times using phosphate buffered saline (PBS). The viability of cells was checked using trypan blue(cell viability should be above 98 %).

The cell suspension was added to tubes containing various concentrations of the test compounds, and the volume was made up to 1 mL using PBS. Control tubes containing only cell suspension. These assay mixtures were incubated for 3 h at 37 °C, and then 1 mL of trypan blue was added after incubation, and the number of dead cells were counted using a haemocytometer³⁶. The percentage cytotoxicity was calculated using the following equation.

% Cytotoxicity =
$$\frac{No. of dead cell}{No. of viable cell + No. of dead cells} \times 100$$

MTT assay

The ability of the cells to survive a toxic insult is the basis of most cytotoxic assays. This assay is based on the assumption that dead cells or their products do not reduce tetrazolium. The assay depends both on the mitochondrial activity per cell and number of cells present. The cleavage of 3-(4,5-dimethylthiazole-2yl); 2, 5-diphenyl tetrazolium bromide (MTT) to a blue formazan derivative by living cells is a very effective principle on which the assay is based. The principle involved is the cleavage of tetrazolium salt, 3-(4, 5-dimethyl thiazole-2yl); 2, 5diphenyl tetrazolium bromide into a blue coloured product (formazan) by mitochondrial enzyme succinate dehydrogenase. The number of cells was found to be proportional to the extent of formazan production by the cells used³⁷. Cells were seeded in a 96-well flat-bottom plate (5000 cells/well) and permitted to adhere for 24 h at 37 $^{\circ}$ C with 5 % CO₂ atmosphere. Different drug concentration was added and incubated further for 48 hrs. Before 4 h of the completion of incubation, 20 µL of MTT (5 mg/mL) was added. Dead cell percentage was determined using an ELISA plate reader set to record absorbance at 570 nm. The percentage growth inhibition was calculated using the formula given below³⁸.

% Growth inhibition =
$$100 \frac{OD \text{ of individual test group}}{OD \text{ of control group}} \times 100$$

Results

Extract yield and Preliminary qualitative phytochemical analysis

The soxhlet extraction of *P. crassicaulis* Bremek. leaf (750 g) with petroleum ether gives 12.32 g, with chloroform gives 19.99 g, with ethanol gives 75.35 g and with water gives 45 g yield.

The qualitative phytochemical analysis of *P. crassicaulis* Bremek. leaf extracts revealed that leaf petroleum ether and chloroform extracts showed nil phytoconstituents and leaf ethanolic extract confirms the presence of saponins, tannins, flavonoids, steroids/sterols, glycosides and phenols and in aqueous extract confirms the presence of saponins and glycosides (Table 1).

	alitative preliminary phytochem	-			
Secondary Metabolites	Type of tests	Petroleum ether crude extract	Chloroform crude extract	Ethanolic crude extract	Aqueous crude extract
		-	-	Leaf	Leaf
Alkaloids	Mayer's test	-	-	-	-
	Wagner's test	-	-	-	-
Saponins	Foam test	-	-	+	+
Tannins	Ferric chloride test	-	-	+	-
Flavonoids	Shinda test	-	-	+	-
	Zinc -HCl reduction test	-	-	+	-
	Alkaline reagent test	-	-	+	-
	Lead acetate test	-	-	+	-
Steroids	Salkowaski test	-	-	+	-
Glycosides	Keller-Killianis test	-	-	+	+
-	Brown water test	-	-	+	-
	Legal test	-	-	+	-
Phenols	Ferric chloride test	-	-	+	-
	Acetic acid test	-	-	+	-
Sterols	Liebermann burchad test	-	-	+	-
+: positive result, -: Neg	ative result				

Quantitative phenol and flavonoid analysis

Total phenolic content in leaf ethanolic extract was expressed as equivalent to gallic acid (EGA) and was found to be 654.3 ± 0.32 of dry sample, respectively. The total flavonoid content of the sample was found 385.1 ± 0.95 of dry extract as equivalent to quercetin in samples respectively.

GC-MS analysis of leaf ethanolic crude extract

In GC-MS analysis of medicinal P. crassicaulis Bremek. ethanolic leaf extract revealed the presence of 36 compounds, but peak 5 is undetected and peak 33 and 34 is the same compound in different quantity, in that major percentage compounds of present were 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl), Phenol (38.84 %), Cyclo{tetra[(5-tbutyl-2 hydroxy-1,3-phenylene)methylene]} (15.72 %), 4H-Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl-(9.34 %), Bicyclo[3.3.1]nona-3,7-diene-2,9-dione (6.25 %), Methyl Ester Of Bicyclo[4.3.0]Non-1(6)-En-4,7-Dione-8-Carboxylic acid (5.66 %). The compounds such 2-Tert-Butyl-4, 6-Bis (3, 5-Di-Tert-Butyl-4as Hydroxybenzyl) Phenol (37.44 %) has anti-oxidant and

UV stabilizing properties. Cyclo {tetra[(5-t-butyl-2-hydroxy-1,3-phenylene)methylene]} (15.72 %), 4H-Pyran-4-one and 2,3-dihydro-3,5-dihydroxy-6-methyl-(9.34 %) has antioxidant properties (Table 2; Fig. 1 and 2).

In vitro antioxidant properties

In the all the experiments leaf aqueous extract showed less anti-oxidant activity compared to the leaf ethanolic crude extracts, so in the following, only the leaf ethanolic extract was compared with pure compound and standard.

Total antioxidant and reductive capability

The total antioxidant activity of both ethanolic crude extract and the pure compound had shown significant antioxidant activity. The pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol activity was comparable to that of standard ascorbic acid.

However, ethanolic crude extract and pure compound have shown appreciable reductive capability when compared to standard quercetin. Like antioxidant activity, the reducing power of ethanolic crude extract and pure compound were increased with

Table 2 — Presence of metabolites in GC-MS analysis of crude ethanolic extract of <i>Pavetta crassicaulis</i> Bremek. leaf collected from	
Western Ghats Karnataka	

S. No.	Chemical compound present	Average percentage	Properties of the compound
1	2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3-one	0.53	Food-grade flavor ingredients ³⁹
2	2,5-Dimethyl-4-hydroxy-3(2H)-furanone	0.52	Flavor and perfume industry ⁴⁰
3	2-Hexanone, 3-methyl-4-methylene-	1.45	Paint and paint thinner ⁴¹
4	4H-Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl-	9.34	Mutagen Antimicrobial, anti-inflammatory and antioxidant capacity ⁴²⁻⁴⁴
5	4-[4-Chlorophenyl]-N-[2-[1-methyl-2- pyrrolidinyl]ethyl]-6-[trichloromethyl]-2-pyrimidine	1.14	Unknown
6	2,3-Dihydro-Benzofuran -	1.35	Entactogen drug of the phenethylamine and amphetamine classes, cytotoxic ⁴⁵
7	2-Furancarboxaldehyde, 5-(hydroxymethyl)-	0.66	Food additives, Antimicrobial, Preservative, Flavoring Agents ⁴⁶⁻⁴⁷
8	1,2,3-Propanetriol, 1-acetate	1.46	Unknown
9	6-Oxoheptanoic acid	1.53	6-Oxoheptanoic acid is a reagent to synthesise new penicillin containing keto acids as side chains. It is also used to study the various metabolic pathways of 4-hydroxypentanoate and Levulinate ⁴⁸
10	Benzaldehyde, 4-hydroxy-	0.47	flavour and fragrance agents ⁴⁹
11	2-Methoxy-4-vinylphenol	1.14	Flavoring agent, antibacterial activity, anti-inflammatory $effect^{50-52}$
12	Phenol, 2-methoxy-4-(2-propenyl)-	0.39	flavoring agent used in the manufacture of vanillin, Anti-Infective Agents, Anti-oxidant ⁵³⁻⁵⁴
13	2,4-Dimethyl-3-nitrobicyclo[3.2.1]octan-8-one	0.78	Oils obtained from myrrh and frankincense and parthenium weed have little percentage ⁵⁵⁻⁵⁶
14	rac-2,4-Dimethyl-3-nitrobicyclo[3.2.1]octan-8-one	0.96	Unknown
15	Benzaldehyde, 2-hydroxy-6-methyl-	0.98	Pheromone of the acarid mite <i>Tyrophagus pernicious</i> and Grain Mite <i>Aleuroglyphus ovatus</i> ⁵⁷⁻⁵⁸

~	Western Gha		
S. No.	Chemical compound present	Average Percentage	Properties of the compound
16	2(3H)-Naphthalenone, 4,4a,5,6-tetrahydro-	0.77	Unknown
17	1,5-Diazocine, octahydro-1,5-dinitro-	0.40	Unknown
18	Methyl Ester Of Bicyclo[4.3.0]Non-1(6)-En-4,7-Dione- 8-Carboxylic acid	5.66	Unknown
19	Acetic acid, (2-isopropenyl cyclopentylidene)-, methyl ester	0.83	Unknown
20	Bicyclo[3.3.1]nona-3,7-diene-2,9-dione	6.25	Unknown
21	4-((1E)-3-Hydroxy-1-propenyl)-2-methoxyphenol	1.32	Antimicrobial, Antioxidant, Anti-inflammatory, Analgesic ⁴⁶
22	2-Methyl-5-(4-methylphenyl)tetrazole	0.71	Unknown
	2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*- (E)]]-	1.12	Antimicrobial, anticancer, anti-inflammatory, diuretic, Cytotoxic, Flavoring Agents, used in the preparation of vitamins E and K1. It is also a decomposition product of chlorophyll, used in the treatment of arthritis ^{59-60, 43}
	Hexadecanoic acid, methyl ester	0.29	Perfumes and Cosmetics ⁶¹
	Hexadecanoic acid	2.04	Perfumes, Cosmetics, Enzyme Inhibitors, Surfactants, Flavoring Agents, Adhesives and sealant chemicals, Agricultural chemicals (non-pesticidal), Fillers, Finishing agents, Intermediates, Lubricants and lubricant additives, Surface active agents, antiandrogenic flavour, hemolytic, 5- Alpha-reductase inhibitor ⁶²⁻⁶³
	Octanal, 7-methoxy-3,7-dimethyl-	0.37	Unknown
27	9,12,15-Octadecatrienoic acid, methyl ester, (Z,Z,Z)-	0.28	Anti-bacterial, Anti-inflammatory, hypocholesterolemic, cancer preventive, hepatoprotective, nematicide, insectifuge, antihistaminic, antieczemic, antiacne, 5-Alpha reductase inhibitor, anti-androgenic, anti-arthritic, anti- coronary, insectifuge ⁴³
28	Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester	0.62	Wound healing activity, Hemolytic, pesticide, flavour, antioxidant ⁶⁴
29	9,12-Octadecadienoic acid (Z,Z)-, 2-hydroxy-1- (hydroxymethyl)ethyl ester	0.30	Hypocholesterolemic, Nematicide, Anti-arthritic, Hepatoprotective, Anti- androgenic,Hypocholesterolemic, Nematicide, 5-Alpha reductase inhibitor, Anti- histaminic,Anticoronary Insectifuge,Anti-eczemic,Anti- acne ⁶⁵
30	2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23- hexamethyl-	0.52	Bactericide, Antifungal, Cytotoxic, Antibacterial, Antioxidant, Antitumor, Cancer preventive, Immunostimulant, Chemopreventive, Lipoxygenase- inhibitor, Perfumery, Pesticide and Sunscreen ⁶⁶
31	2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4- Hydroxybenzyl)Phenol	38.84	Unknown
32	Cyclo {tetra[(5-t-butyl-2-hydroxy-1,3- phenylene)methylene]}	15.72	Unknown
33	Stigmast-5-en-3-ol, (3.beta.,24S)-	0.95	Antimicrobial antioxidant, Anti-inflammatory anti-arthritic, anti-asthma, diuretic ⁶⁷

Table 2 — Presence of metabolites in GC-MS analysis of crude ethanolic extract of *Pavetta crassicaulis* Bremek. leaf collected from Western Ghats Karnataka — (*Contd.*)

increasing concentration. Hence the pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxy benzyl)Phenol, can serve as free radical inhibitors or scavengers, which are much capable as standard used (Fig. 3-6).

DPPH radical scavenging activity

DPPH radical scavenging activity was expressed in terms of scavenging percentage and IC_{50} values

(μ g/mL). By the experiment, it is revealed that leaf aqueous crude extracts show negligible scavenging activity compared with leaf ethanolic extract in all the tested concentrations. Compared the crude extracts, leaf (IC₅₀:71.09±0.39 µg/mL) ethanolic crude extract showed appreciable activity compared to the leaf aqueous (IC₅₀: 113.68±1.48 µg/mL) extract. The pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol (IC₅₀: 46.14±1.50 µg/mL)

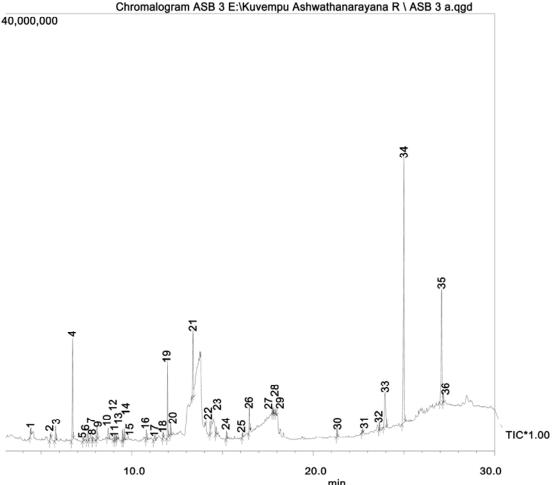


Fig. 1 — GC-MS chromatogram of Pavetta crassicaulis Bremek leaf ethanolic extract

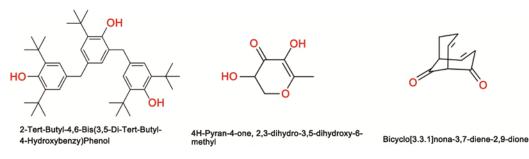


Fig. 2 — Major compounds present in ethanolic extract of Pavetta crassicaulis Bremek. leaf and their structure

displayed appreciable radical scavenging activity which is almost comparable to the standard Ascorbic acid (IC₅₀: DPPH: 39.48±0.02 µg/mL) used (Table 3 and Table 8).

ABTS free radical scavenging activity

ABTS radical scavenging activity was expressed in terms of percentage of inhibition and IC₅₀: values (μ g/mL). Results revealed that leaf ethanolic crude extract shows excellent ABTS radical scavenging activity compared with the aqueous crude leaf extract. The leaf ethanolic extract (IC₅₀:171.5±3.50 µg/mL) demonstrated appreciable ABTS radical scavenging activity compared to the leaf aqueous extract (278.96 \pm 1.50 µg/mL). The pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol which is extracted from leaf ethanolic extract showed excellent radical scavenging activity (IC₅₀: ABTS: 88.14±0.56 µg/mL,) showed excellent activity which is comparable with the standard

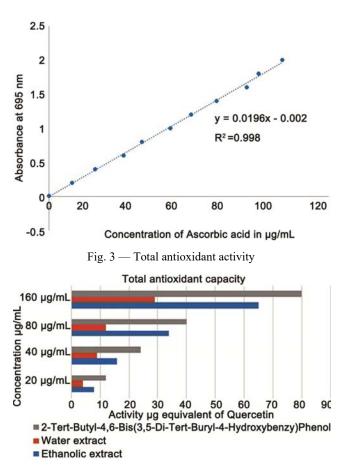


Fig. 4 — Total antioxidant properties of ethanol, water extract with pure compound

Butylated Hydroxylanisole (IC₅₀: ABTS: 66.92 ± 0.36 µg/mL) (Table 3 and Table 8).

Metal chelating activity

In metal chelating activity experiment leaf ethanolic crude extracts showed (IC₅₀:448.36 \pm 1.50 µg/mL) appreciable metal chelating activity compared to the leaf aqueous crude extracts (IC₅₀:1441.87 \pm 1.15 µg/mL). The pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol,

showed excellent metal chelating activity (IC₅₀: $307.48\pm3.45 \ \mu g/mL$) which is lesser value than the standard EDTA (IC₅₀:213.69\pm0.32 \ \mu g/mL) used (Table 4 and Table 10).

Superoxide radical scavenging activity (NBT)

In Superoxide radicals scavenging NBT reduction method, both leaf and flower aqueous crude extracts shows moderate activity but compared to the aqueous extract (IC₅₀: 396.44 \pm 2.69 µg/mL) to leaf ethanolic crude extract (IC₅₀:227.42 \pm 1.32 µg/mL) showed appreciable radical scavenging activity. The pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-

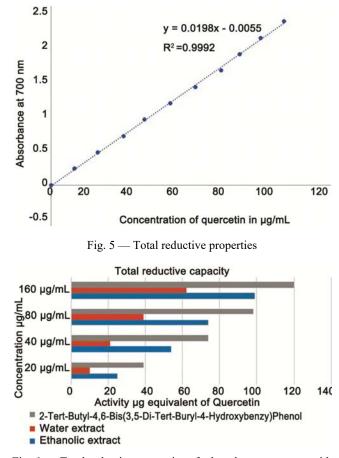


Fig. 6 — Total reductive properties of ethanol, water extract with pure compound

Hydroxybenzyl)Phenol, showed excellent radical scavenging activity (IC50: $148.09\pm0.50 \ \mu g/mL$) compared to standard Gallic acid (IC₅₀:102.17±0.49 $\mu g/mL$) (Table 4 and Table 9).

Hydroxyl radical scavenging activity

In this experiment all the crude extracts showed appreciable activity compared to leaf aqueous extract (IC₅₀: $623.67\pm2.20 \ \mu g/mL$) the leaf ethanolic extract (IC₅₀: $284.42\pm2.50 \ \mu g/mL$) showed excellent scavenging activity.The pure compound "2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-

Hydroxybenzyl)Phenol" shows excellent activity (IC₅₀: 213.48 \pm 1.45 µg/mL) nearer to the value of the standard Gallic acid (IC₅₀: 200.51 \pm 2.45 µg/mL) used (Table 5 and Table 9).

In vitro cytotoxic properties

Experiment was conducting using DLA and EAC cancer cell lines. The aqueous extract was not used because the leaf ethanolic extract in preliminary phytochemical analysis showed positive confirmation results for almost all secondary metabolites and also

Activity	Plant part	Extracts	Concentration in µg/mL	scavenging activity	IC ₅₀ value	Standard Ascorbic acid scavenging activity	IC ₅₀ value of Standard Ascorbic acid
		Aqueous	25	22.54±0.2		76.23±0.23	
			50	34.52±0.05		82.32±0.43	
			75	45.54±0.2		113.11±0.09	
			100	50.2 ± 0.1	113.68±1.48	134.54 ± 0.91	
			125	54.5 ± 0.05	113.08±1.48	156.43 ± 0.02	
			150	60.66 ± 0.05		176.65 ± 0.34	
			175	$62.34{\pm}0.1$		189.41 ± 0.54	
DPPH radical	T £		200	$65.54{\pm}0.4$		210.87±0.32	
scavenging activity	Leaf	Ethanol	25	44.55±0.66		76.23±0.23	39.48 ± 0.02
			50	57.77±0.03		82.32±0.43	
			75	65.48 ± 0.05		113.11±0.09	
			100	77.65 ± 0.65	71.09±0.39	134.54 ± 0.91	
			125	87.32±0.1		156.43 ± 0.02	
			150	92.42±0.66		176.65 ± 0.34	
			175	100.23 ± 0.35		189.41 ± 0.54	
			200	107.54 ± 0.54		210.87±0.32	
Activity	Plant part	Extracts	Concentration in µg/mL	scavenging activity	IC ₅₀ value	Standard µg/mL (Butylated Hydroxyl Anisole)	IC ₅₀ value of Standard Butylated Hydroxyl Anisole
			50	10.23±0.54		47.34±0.32	
			100	18.32 ± 0.12		84.65±0.05	
			150	23.43±0.6		120.43±0.36	
			200	34.22±0.05	278.96±1.50	149.68±0.1	
		Aqueous	250	43.54±0.3	2/8.96±1.50	185.65±0.3	
			300	55.67 ± 0.45		214.76±0.62	
			350	64.87 ± 0.42		254.36±0.06	
ABTS Radical	T £		400	72.34±0.6		287.98 ± 0.6	((02+0.2(
Scavenging activity	Leaf		50	25.43±0.17		$47.34{\pm}0.32$	66.92±0.36
			100	34.56±0.1		84.65±0.05	
			150	49.87±0.32		120.43±0.36	
		Ethanol	200	55.32 ± 0.4	171 5 2 50	149.68 ± 0.1	
		Einanoi	250	67.82 ± 0.34	171.5±3.50	185.65±0.3	
			300	86.11±0.2		214.76±0.62	
			350	$94.54{\pm}0.4$		254.36±0.06	
			400	111.13±0.05		287.98 ± 0.6	

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* IC_{50} value is the amount of extract needed for scavenging 50 % of the radical produced in the reaction mixture. Values are mean \pm standard deviation of three independent replicates.

showed excellent antioxidant properties, so, leaf extract with the pure compound tested for cytotoxic properties using DLA and EAC cells, revealed that the leaf extract showed appreciable cytotoxic properties and pure compound showed appreciable cytotoxic properties but less than the standard used.

Trypan blue dye exclusion technique

In this method leaf ethanolic crude extract and pure compound was tested for its cytotoxicity, revealed that leaf ethanolic crude extract is effective against DLA (CTC₅₀: 283.62 \pm 6.87 µg/mL) and EAC cells (CTC₅₀: 239.53 \pm 6.9 µg/mL) but compared to standard it is negligible. The pure compound 2-Tert-Butyl-4, 6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol showed moderate cytotoxic activity for both DLA (CTC₅₀: 103.48±0.5 μ g/mL) and EAC cells (CTC₅₀: 271.42±3.4 μ g/mL) which is not comparable to the standard curcumin (CTC₅₀: 54.31±1.5 μ g/mL) (Table 6 and 11).

MTT assay

The leaf ethanolic crude extract and pure compound were tested for its cytotoxicity using MTT assay and the leaf ethanolic crude extract showed appreciable cytotoxicity against DLA (CTC_{50} :

	Table 4 — Metal che	-	-	-	-			
S. No	. Activity	Plant part	Extracts	Concentration in µg/mL	scavenging activity	IC ₅₀ value	Standard Ascorbic acid scavenging activity	IC ₅₀ value of Standard EDTA
			Aqueous	200	7.65 ± 0.17		58.76±0.32	
				400	14.34 ± 0.12		98.34±0.03	
				600	21.34 ± 0.06		144.65 ± 0.32	
				800	27.98 ± 0.62	1441.87±1.15	195.76 ± 0.45	
				1000	36.54 ± 0.34	1441.8/±1.13	242.87 ± 0.14	
				1200	$42.34{\pm}0.2$		283.24 ± 0.36	
				1400	49.67 ± 0.4		332.31±0.05	
1	Metal chelating	Leaf		1800	56.75 ± 0.05		375.52 ± 0.82	
	activity		Ethanol	200	37.43 ± 0.14		58.76 ± 0.32	213.69±0.32
				400	55.67 ± 0.03		$98.34{\pm}0.03$	215.07±0.52
				600	71.23 ± 0.6	448.36±1.50	144.65 ± 0.32	
				800	95.43 ± 0.32		195.76±0.45	
				1000	124.32 ± 0.54		242.87 ± 0.14	
				1200	132.45 ± 0.42		283.24 ± 0.36	
				1400	143.26 ± 0.05		332.31±0.05	
				1800	$165.43 {\pm} 0.05$		375.52 ± 0.82	
	Activity	Plant part	Extracts	Concentration in	scavenging	IC50 value	Standard µg/mL	IC50 value of
				μg/mL	activity		(Butylated Hydroxy)	
							Anisole)	acid
				100	19.08 ± 0.2		51.66±0.11	
				200	24.56 ± 0.05		95.67±0.54	
				300	39.34±0.2		148.12 ± 0.42	
			Aqueous	400	50.32 ± 0.1	396.44±2.69	199.77±1.34	
			. queeus	500	62.45±0.05		247.32 ± 0.49	
				600	78.32±0.05		282.22±0.19	
2	Superoxide radicals			700	89.65±0.1		341.21±0.24	
	scavenging NBT	Leaf		800	90.32±0.4		395.74±0.63	102.17 ± 0.49
	reduction method	Loui		100	31.32 ± 0.32		51.66±0.11	102.17±0.1
				200	51.22 ± 0.03		95.67±0.54	
				300	73.23 ± 0.32		148.12 ± 0.42	
			Ethanol	400	87.65 ± 0.45	227.42±1.32	199.77±1.34	
			Lanunol	500	99.43±0.14		247.32 ± 0.49	
				600	123.65 ± 0.36		282.22±0.19	
				700	153.22 ± 0.05		341.21±0.24	
				800	171.76 ± 0.82		395.74±0.63	

 $*IC_{50}$ value is the amount of extract needed for scavenging 50% of the radicals produced in the reaction mixture. Values are mean \pm standard deviation of three independent replicates.

295.39 \pm 8.81 µg/mL) and EAC (CTC₅₀: 267.68 \pm 10.68 µg/mL) cells but not as good as standard. The pure compound "2-Tert-Butyl-4, 6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol" showed excellent cytotoxic activity DLA (CTC₅₀: 117.91 \pm 1.5 µg/mL) and EAC cells (CTC₅₀: 251.68 \pm 2.5 µg/mL) which is not comparable to the standard curcumin (CTC₅₀: 54.31 \pm 1.5 µg/mL) (Table 7 and 12).

Discussion

Preliminary qualitative phytochemical analysis

Preliminary quantitative phytochemical analysis of *P. crassicaulis* Bremek. petroleum ether, chloroform crude extracts showed negative results for all the tested

phytochemicals, aqueous extract gives positive results for saponins and glycosides, but the ethanolic crude extract gives positive confirmation test for saponins, tannins, flavonoids, steroids/sterols, glycosides and phenols. This test confirms that the ethanol dissolves all the phytochemicals from *P. crassicaulis* Bremek. in a sufficient quantity to influence different phytochemical and pharmacological activities.

GC-MS analysis

Preliminary quantitative phytochemical analysis of *P. crassicaulis* Bremek. crude extracts give a clear picture of phytochemical dissolved in leaf ethanolic extract so, we subjected only ethanolic crude extract

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	Table	5 — Hydroxy	l radical scavenging	assay of Pavetta cra.	ssicaulis Brem	ek. leaf extracts	
Activity	Plant part	Extracts	Concentration in µg/mL	scavenging activity	$\rm IC_{50}$ value	Standard EDTA scavenging activity	IC ₅₀ value Standard EDTA
			100	9.65±0.45		32.14±0.63	
			200	$16.34{\pm}0.14$		58.76±0.32	
			300	$22.34{\pm}0.2$		76.58 ± 0.98	
		Aqueous	400	$28.98{\pm}0.05$	623.67±2.20	98.34±0.03	
			500	$37.54{\pm}0.1$		119.24±0.19	
			600	49.34±0.4		144.65±0.31	
			700	55.67±0.61		172.23 ± 0.48	
Hydroxyl radical	Leaf		800	68.75±0.34		195.76±0.45	
scavenging assay	Lear		100	26.43 ± 0.32		32.14±0.63	200.51±2.45
			200	$40.32{\pm}0.05$		58.76±0.32	
			300	56.43±0.36		76.58 ± 0.98	
		Ethanol	400	79.11±0.1	284.42±2.50	98.34±0.03	
			500	87.54±0.3		119.24±0.19	
			600	99.26±0.62		144.65±0.31	
			700	118.34±0.06		172.23±0.48	
			800	125.43±0.6		195.76±0.45	

 $*IC_{50}$ value is the amount of extract needed for scavenging 50 % of the radicals produced in the reaction mixture. Values are mean± standard deviation of three independent replicates

Table 6 — <i>In-vitro</i> cytotoxic screening by <i>Pavetta crassicaulis</i> Bremek. leaf ethanolic extract against DLA and EAC cells by Trypan blue
dye exclusion technique

S. No.	Concentration (µg/mL)	Leaf extract against DLA cells		Leaf extract against I	Standard	control	
		Percentage cytotoxicity	CTC ₅₀	Percentage cytotoxicity	CTC ₅₀	(Curcumin)	
1	10	$0{\pm}0$		1 ± 1		15.4±3.3	
2	20	8.33±1.15		8.66 ± 1.52		34.4 ± 3.3	
3	50	12.66 ± 2.08	$283.62{\pm}6.87$	15±1.73	239.53 ± 6.9	100 ± 0.5	0.23 ± 1.54
4	100	17±2.64		23.33 ± 2.08		100 ± 0.5	
5	200	29±1		31.33±0.57		100 ± 0.5	

Table 7 — In-vitro cytotoxic screening by Pavetta crassicaulis Bremek. leaf ethanolic extract against DLA and EAC cells by MTT assay

S.	Concentration	Leaf extract against DLA cells			Leaf ext	ract against l	EAC cells	Absorbance of	Standard	control
No.	(µg/mL)	Absorbance	Percentage	CTC ₅₀	Absorbance	Percentage	CTC ₅₀	standard	(Curcumin) %	
		at 570 nm	cytotoxicity		at 570 nm	cytotoxicity		Curcumin at	cytotoxicity	
								570 nm		
1	10	8±1.52	0.211 ± 0.24		$7.97{\pm}2.08$	0.33 ± 0.57		0.211 ± 0.24	15.4±3.3	
2	20	7.33 ± 0.57	$0.123{\pm}0.18$		7.41 ± 3.51	7.33 ± 1.52		0.123 ± 0.18	34.4 ± 3.3	
3	50	$7.04{\pm}1.54$	$0.001{\pm}0.01$	$295.39{\pm}8.81$	$6.88{\pm}0.5$	14 ± 3	267.68±10.68	$0.001{\pm}0.01$	100 ± 0.5	$0.29{\pm}0.5$
4	100	6.77 ± 0.50	$0.001{\pm}0.01$		6.35 ± 1.52	20.66 ± 3.51		$0.001 {\pm} 0.01$	100 ± 0.5	
5	200	5.79 ± 1.50	$0.001{\pm}0.01$		5.71 ± 0.57	28.66 ± 2.08		$0.001 {\pm} 0.01$	100 ± 0.5	

for GC-MS revealed the presence of 33 compounds (Table 2). Among all the confirmed phytochemicals 21 compounds reported for many medicinal and pharmacological properties and 12 compounds activity was not reported. Among all the phytochemicals present in the GC-MS analysis 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol (38.84) was the major compound, followed by Cyclo{tetra[(5-t-butyl-2hydroxy-1,3-phenylene)methylene]}(15.72 %) not reported for its pharmacological properties. The next successive compound 4H-Pyran-4-one, 2,3-dihydro-3,5dihydroxy-6-methyl- (9.3 4 %) was reported for many pharmacological properties including antioxidant properties⁴²⁻⁴⁴.

All the other phytocompounds like Bicyclo[3.3.1]nona-3,7-diene-2,9-dione (6.25 %), Methyl Ester of Bicyclo[4.3.0]Non-1(6)-En-4,7-Dione-8 (5.66 %), Hexadecanoic acid (2.04%), 6-Oxoheptanoic acid (1.53 %), 1,2,3-Propanetriol, 1-acetate (1.46 %), 2-Hexanone, 3-methyl-4-methylene (1.45 %), 2,3-Dihydro-Benzofuran (1.35 %), 4-((1E)-3-Hydroxy-1-propenyl)-2methoxyphenol (1.32 %), 4-[4-Chlorophenyl]-N-[2-[1methyl-2-pyrrolidinyl]ethyl]-6-[trichloromethyl]-2pyrimidine (1.14%), 2-Methoxy-4-vinylphenol (1.14%), 2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)] (1.12 %), Benzaldehyde, 2-hydroxy-6-methyl (0.98 %),

		Pavetta crassicaulis	Bremek. leaf ext	ract		
S. No. Test sample	Activity	Concentration in µg/mL	Scavenging activity	IC ₅₀ value	Standard ascorbic acid scavenging activity	IC ₅₀ value of standard ascorbic acid
		25	33.21±0.43		76.23±0.23	
		50	55.32 ± 0.33		82.32±0.43	
	DPPH	75	11.42 ± 0.32		113.11 ± 0.09	
1	radical scavenging	100	132.43 ± 2.08	46.14±1.50	134.54 ± 0.91	
1	activity	125	155.31 ± 3.51	40.14±1.30	156.43 ± 0.02	39.48±0.02
	activity	150	164.92 ± 3		176.65 ± 0.34	
		175	189.23 ± 1.52		189.41 ± 0.54	
		200	233.44 ± 0.57		210.87 ± 0.32	
2-Tert-Butyl-4,6- Bis(3,5-Di-Tert-Butyl-4- Hydroxybenzyl)Phenol		Concentration in µg/ml	Scavenging activity	IC50 value	Standard Butylated Hydroxylanisole scavenging activity	IC ₅₀ value of standard Butylated Hydroxylanisole
		50	33.45±0.23		47.34±0.32	
		100	57.54 ± 0.43		84.65±0.05	
	ABTS	150	83.43 ± 0.09		120.43 ± 0.36	
2	Radical	200	$100.32{\pm}0.91$	88.14±0.56	149.68 ± 0.1	66.92±0.36
2	Scavengin	250	123.65 ± 0.02	88.14±0.30	185.65±0.3	
	g activity	300	167.86 ± 0.34		214.76±0.62	
		350	198.4 ± 0.54		254.36 ± 0.06	
		400	256.43 ± 0.32		$287.98 {\pm} 0.6$	

Table 8 — DPPH and ABTS assay of pure compound "2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol" extracted from
2-ref-buty-4,0-bis(5,5-bi-ref-buty-4-fryutoxybenzyf) inclusion extracted from
Pavetta crassicaulis Bremek, leaf extract

 $*IC_{50}$ value is the amount of extract needed for scavenging 50 % of the radical produced in the reaction mixture. Values are mean \pm standard deviation of three independent replicates. ND- not determined

 Table 9 — NBT and Hydroxyl radical scavenging assay of pure compound "2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol" extracted from *Pavetta crassicaulis* Bremek. leaf extract

S. No.	Test sample	Activity	Concentration in µg/mL	Scavenging activity	IC_{50} value	Standard Gallic acid scavenging activity	IC ₅₀ value of standard Gallic acid
			100	31.26±0.63		51.66±0.11	
			200 65.43±0.32		95.67±0.54		
		Superoxide radicals scavenging NBT reduction method	300	89.43±0.98	148.09±0.54	148.12 ± 0.42	
			400	134.23±0.03		199.77±1.34	
I			500	154.33±0.19		247.32±0.49	102.17 ± 0.49
			600	198.59±0.31		282.22±0.19	
	2-Tert-		700	243.54 ± 0.48		341.21±0.24	
	Butyl-4,6-		800	298.6±0.45		$395.74{\pm}0.63$	
	Bis(3,5-Di-		Concentration	Scavenging		Standard EDTA	IC_{50} value of standard
	Tert- Butyl-4-		in µg/mL	activity	IC ₅₀ value	scavenging activity	EDTA
	Hydroxyben		100	35.23±0.42		32.14±0.63	
	zyl)Phenol	Hydroxyl radical scavenging assay	200	66.43±0.19		58.76 ± 0.32	
			300	96.23±0.63		76.58 ± 0.98	
2			400	123.12 ± 0.98	213.48±1.45	98.34 ± 0.03	
			500	124.91 ± 0.03	213.40±1.43	119.24 ± 0.19	200.51±2.45
			600	150.43 ± 0.27		144.65 ± 0.31	
			700 182.33 ± 0.76			172.23 ± 0.48	
			800	210.87 ± 0.03		195.76±0.45	

 $*IC_{50}$ value is the amount of extract needed for scavenging 50 % of the radical produced in the reaction mixture. Values are mean \pm standard deviation of three independent replicates. ND- not determined

	8 7 1	Pavetta cra	ssicaulis Bremek	. leaf extract	5 5 5	,
S. No. Test sample	Activity	Concentrati on in µg/mL	Scavenging activity	IC_{50} value	Standard scavenging activity (EDTA)	IC_{50} value of standard EDTA
1 2-Tert-Butyl-4,6- Bis (3,5-Di-Tert- Butyl-4- Hydroxybenzyl)Ph enol	Metal chelating assay	200 400 600 800 1000 1200 1400 1800	34.23 ± 0.32 66.54 ± 0.54 95.47 ± 1.54 120.76 ± 1.54 165.48 ± 2.5 198.76 ± 3.4 234.54 ± 0.43 287.54 ± 0.23	307.48±2.43	58.76 ± 0.32 98.34 ± 0.03 144.65 ± 0.32 195.76 ± 0.45 242.87 ± 0.14 283.24 ± 0.36 332.31 ± 0.05 375.52 ± 0.82	213.69±0.32

Table 10 — Metal chelating assay of pure compound "2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol" extracted from

 $*IC_{50}$ value is the amount of extract needed for scavenging 50 % of the radical produced in the reaction mixture. Values are mean \pm standard deviation of three independent replicates. ND- not determined

Table 11 — Trypan blue dye exclusion technique "2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol" extracted from *Pavetta crassicaulis* Bremek. leaf extract

S. No.	Concentration (µg/mL)	Trypan blue dye exclusion technique for DLA cells		Trypan blue dye exclusion technique for EAC cells		Standard	Standard	Control
		Percentage cytotoxicity	CTC ₅₀	Percentage cytotoxicity	CTC ₅₀	(Curcumin)	CTC ₅₀	Control
1	10	5.1±0.5		$0{\pm}0$		15.4±3.3		
2	20	11.3 ± 1.5		$0{\pm}0$		34.4±3.3		
3	50	22.3±0.5	$117.91{\pm}1.5$	10.5 ± 2.2	251.68 ± 2.5	100 ± 0.5	54.31±1.5	0.23 ± 1.54
4	100	44.55±2.5		21.45±3.4		100 ± 0.5		
5	200	77.88±1.5		43.54±1.5		$100{\pm}0.5$		

Table 12 — MTT assay of pure compound "2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol" extracted from Pavetta crassicaulis Bremek. leaf extract

S. Concentration MTT assay for DLA cells			MTT a	assay for EA	C cells	Absorbance	Standard	CTC50 of Control		
No.	(µg/mL)	Absorbance	Percentage	CTC ₅₀	Absorbance Percentage CTC ₅₀			of standard Curcumin % standard		
		at 570 nm	cytotoxicity		at 570 nm	cytotoxicity		Curcumin	cytotoxicity	(Curcumin)
								at 570 nm		
1	10	7.34±1.5	8±1.5		8.10 ± 3.4	$0{\pm}0$		0.211 ± 0.24	15.4 ± 3.3	
2	20	6.69 ± 2.5	16.43 ± 2.3		8±1.3	0 ± 0		$0.123{\pm}0.18$	34.4 ± 3.3	
3	50	5.18 ± 0.5	35.21±1.2	103.48±0.5	$7.60{\pm}2.5$	5±1.4	271.42±3.4	$40.001{\pm}0.01$	100 ± 0.5	54.31±1.5 0.31±1.5
4	100	3.41 ± 2.5	57.43±2.3		$6.24{\pm}0.01$	22±1.5		$0.001{\pm}0.01$	100 ± 0.5	
5	200	2.68 ± 0.01	66.54±2.3		$4.56{\pm}0.01$	43±1.5		$0.001 {\pm} 0.01$	100 ± 0.5	

rac-2,4-Dimethyl-3-nitrobicyclo[3.2.1]octan-8-one (0.96 %), Stigmast-5-en-3-ol, (3.beta.,24S) (0.95%), Acetic acid, (2-isopropenyl cyclopentylidene)-, methyl ester (0.83 %), 2,4-Dimethyl-3-nitrobicyclo[3.2.1]octan-8-one (0.78 %),2(3H)-Naphthalenone, 4,4a,5,6-tetrahydro (0.77 %), 2-Methyl-5-(4-methylphenyl)tetrazole (0.71 %), 2-Furancarboxaldehyde, 5-(hydroxymethyl) (0.66 %), Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester (0.62), 2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3one (0.53 %), 2,5-Dimethyl-4-hydroxy-3(2H)-furanone (0.52)%), 2,6,10,14,18,22-Tetracosahexaene, 2,6,10,15,19,23-hexamethyl (0.52 %), Benzaldehyde, 4hydroxy- (0.47 %), 1,5-Diazocine, octahydro-1,5dinitro- (0.4 %), Phenol, 2-methoxy-4-(2-propenyl) (0.39 %), Octanal, 7-methoxy-3,7-dimethyl (0.37 %),

9,12-Octadecadienoic acid (Z,Z)-, 2-hydroxy-1 (0.3 %), Hexadecanoic acid, methyl ester (0.29 %), 9,12,15-Octadecatrienoic acid, methyl ester, (Z,Z,Z) (0.28 %) were present in a meager percentage have reported for many medical as well as pharmacological properties (Table 2).

The antioxidant experiments like metal chelating activity, superoxide radicals, hydroxyl radical, DPPH radicals, ABTS radical scavenging assays results concluded that leaf ethanolic extract and its extracted pure compound has appreciable antioxidant properties but not good as standards used.

Aqueous extract showed negligible activity which is indirectly confirmed in the qualitative phytochemical analysis only gives the positive results for saponins and glycosides, so its negligible activity was also expected one.

The GC-MS analysis of P. crassicaulis Bremek. ethanolic leaf crude extracts revealed the presence of 33 compounds, and it is evident that phenolic compounds in the plants will contribute to the antioxidant properties. The antioxidant activity of phenolic compounds is mainly due to their redox phenomenon, due to this, they play the role of adsorbing and neutralising free radicals, quenching singlet and triplet oxygen, or decomposing peroxides⁶⁸⁻⁶⁹. GC-MS analysis report of *P. crassicaulis* Bremek. leaf ethanolic extract clearly reveal the presence of phenolic substances like 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl) Phenol (38.84 %), Cyclo{tetra[(5-t-butyl-2-hydroxy-1,3-phenylene) methylene]}(15.72 %), etc. may influence the antioxidant properties of ethanolic extract.

Thus, indicates that the polyphenols present in leaf ethanolic extract could be responsible for the positive effects which in turn give the positive feedback on the folklore use and withstand the use of pavetta plant parts by the tribal folklore medicinal practitioners in local India.

In-vitro cytotoxic experiments like Trypan blue dye exclusion technique and MTT assay against DLA and EAC cancer cells revealed that leaf ethanolic extract showed moderate cytotoxicity and the pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol showed appreciable cytotoxic activity but less than the standard curcumin used (Table 11 and Table 12).

In the crude ethanolic extract of *P. crassicaulis* Bremek. the major 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl)Phenol may influence the cytotoxicity, or all the 33 compounds may also influence cytotoxic activity.

Conclusion

The present study shows that *P. crassicaulis* Bremek. leaf ethanolic crude extract has appreciable antioxidant and cytotoxic properties and its pure compound 2-Tert-Butyl-4,6-Bis(3,5-Di-Tert-Butyl-4-Hydroxybenzyl). Phenol has appreciable antioxidant and cytotoxic properties but not good as the standards used. These results confirm positive activity of the plant as a therapeutic agent in tribal medicine. Thus *P. crassicaulis l*eaf could be exploited as a valuable source of antioxidant and cytotoxic agent for the pharmaceutical industry.

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Reference

- 1 Valko M, Morris H and Cronin M T, Metal toxicity and oxidative stress, *Curr Med Chem*, 2005, **12**, 1161-208.
- 2 Clarkson B D and Burchenal J H, Preliminary screening of antineoplastic drugs, *Prog Clin Cancer*, 1965, 1, 625-9.
- 3 Prasad S B and Giri A, Antitumor effect of cisplatin against murine ascites Dalton's lymphoma, *Indian J Exp Biol*, 1994, 32, 155-62.
- 4 Fenninger L D and Mider G B, Energy and nitrogen metabolism in cancer, *Adv Cancer Res*, 1954, **2**, 229-253.
- 5 Tausz M, Wonisch A, Grill D, Morales D and Jimenez M S, Measuring antioxidants in tree species in the natural environment: from sampling to data evaluation, *J Exp Bot*, 2003, 54,1505.
- 6 Cai Y, Luo and Sun M, Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer, *Life Sci*, 2004, 74, 2157-2184.
- 7 Cowan M M, Plant products as antimicrobial agents, *Clin Microbiol Rev*, 1999, **12**(4), 564-582.
- 8 Demain A L and Sanchez S, Microbial drug discovery: 80 years of progress, *J Antibiot*, 2009, 62(1), 5-16.
- 9 Davies J and Davies D, Origins and evolutions of antibiotic resistance, *Microbiol Mol Biol Rev*, 2010, 74(3), 417-433.
- 10 Chopra R N, Nayar S L and Chopra I C, Glossary of Indian medicinal plants (Publication and Information Directorate, Council of Scientific and Industrial Research, New Delhi) 1956, 256-257.
- 11 Rabe T and van Staden J, Antibacterial activity of South African plants used for medicinal purposes, *J Ethnopharmacol*, 1997, **56**(1), 81-7.
- 12 Kamboj V P, Herbal medicine, Curr Sci, 2000, 78(1), 35-39.
- 13 Manikandan R and Lakshminarasimhan P, Flowering Plants of Rajiv Gandhi (Nagarahole) National Park, Karnataka, India, *Check List*, 2012, 8(6), 1052–1084.
- 14 Jayant K and Mehta P, A Study of status, distribution and dynamics of private and community forests in Sahyadri-Konkan Corridor of Maharashtra Western Ghats, Technical report submitted to CEPF-ATREE, Wildlife Research and Conservation Society, Pune, 2013, 161-163.
- 15 Bhise M R, Rahangdale S S, Rahangdale S R and Kambhar S V, Floristic Study of Kalbhairavanatha Sacred Grove, Terungan, Ambegaon Taluka, Pune, *Research & Reviews:* J Bot, 2013, 2(1), 17 -24.
- 16 Faude U, Human induced fragment formation and vascular plants species diversity: A case study of Satkosia Gorge Wildlife sanctuary and its surroundings, Orissa, India, Department of Geography of the Rheinische Friedrich-Wilhelms –University, Bonn, Germany, Bonn Foundation for Ecological Security, 2008, 52-64.
- 17 Trivedi P C, Silori C S, Dixit, A M, Gupta L and Mistry N, Medicinal plants utilization and conservation observations on medicinal plant richness and associated conservation issues in district Kachchh, Gujarat, 2nd edn, (Aavishkar Publishers, Distributors Jaipur, India), 2009, 165.
- 18 Singh P and Ali S J, Ethnomedicinal plants of family Rubiaceae of eastern Uttar Pradesh, *Indian J Life Sci*, 2012, 1(2), 83-86.

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- 19 Mishra M K, Biodiversity, traditional knowledge and village ecosystem sustainability, *The Ecoscan*, 2013, **3**, 235-240.
- 20 www.pankajoudhia.com/mpd.pdf
- 21 Kar A, Bora D, Borthakur S K, Goswami N K and Saharia D, Wild edible plant resources used by the Mizos of Mizoram, India, *Kathmandu Uni J Sci Eng Technol*, 2013, 9(1), 106-126.
- 22 Kantamreddi V S S N, Lakshmi Y N and Kasapu V V V S, Preliminary phytochemical analysis of some important Indian plant species, *Int J Pharma Bio Sci*, 2010, 1(4), 351-358.
- 23 Ashwathanarayana R and Naika R, Preliminary phytochemical and antimicrobial properties of Pavetta crassicaulis Bremek. flower extracts collected from Western Ghats, Karnataka, India, J Pharmacogn Phytochem, 2016, 5(3), 256-262.
- 24 De-Castro M D L and Ayuso L E G, Soxhlet extraction of solid materials: an outdated technique with a promising innovative future, *Anal Chim Acta*, 1998, 369, 1-10.
- 25 Singleton V L and Rossi J A, Colorimetry of total phenolics with phosphomolybdic-phos photungstic acid reagent, Am J Enol Vitic, 1965, 16, 144-158.
- 26 Zhishen J, Mengcheng T and Jianming W, The determination of flavonoid content in mulberry and their scavenging effects on superoxide radicals, *Food Chem*, 1999, **64**, 555-9.
- 27 Prieto P, Pineda M and Aguilar M, Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphormolybdenum complex: specific application to the determination of vitamin E, *Anal Biochem*, 1999, **269**, 337-41.
- 28 Oyaizu M, Studies on products of browning reaction: Antioxidative activities of products of browning reaction prepared from glucosamine, *Japanese J Nutri*, 1986, 44, 307-15.
- 29 McCord J M and Fridovich I, Superoxide dismutase, An enzymic function for erythrocuprein (hemocuprein), J Biol Chem, 1969, 44, 6049–6055.
- 30 Kunchandy E and Rao M N A, Oxygen radical scavenging activity of curcumin, *Int J Pharm*, 1990, **58**, 237–240.
- 31 Ohkawa H, Ohishi N and Yagi K, Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction, *Anal Biochem*, 1979, **95**, 351–358.
- 32 Alzoreky N and Nakahara N, Antioxidant activity of edible Yemeni plants evaluated by ferryl myoglobin/ABTS assay, *Food Sci Technol Res*, 2001, 7, 141–144.
- 33 Dinis T C P, Madeira V M C and Almeida L M, Action of phenolic derivatives (acetaminophen, salicylate and 5aminosalicylate) as inhibitors of membrane lipid peroxidation as peroxyl radical scavenging effects, *Chem Pharm Bull*, 1994, 36, 2090-7.
- 34 Moldeus P, Hogberg J, Orrhenius S and Parker F S, Methods in Enzymology, (Academic Press, New York), 1978, 52, 60–71.
- 35 Kuttan R, Bhanumathy P, Nirmala K, and George M C, Possible anticancer activity of turmeric, *Cancer lett*, 1985, 29, 197-202.
- 36 Shrivastava S and Ganesh N, Tumor inhibition and cytotoxicity assay by aqueous extract of onion (*Allium cepa*) & Garlic (*Allium sativum*): An *in-vitro* analysis, *Int J phyto Med*, 2010, 2, 80-84.
- 37 Ramnath V, Rekha P S, Kuttan G and Kuttan R, Regulation of Caspase-3 and Bcl-2 expression in Dalton's lymphoma ascites cells by Abrin, *Evid Based Complement Alternat Med*, 2009, 6, 233-238.
- 38 Hajighasemi F and Mirshafiey A, Propranolol effect on proliferation and vascular endothelial growth factor secretion

in human immune competent cells, J Clin Immunol Immunopathol Res, 2010, 2, 22-27.

- 39 Pubchem-2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3-one, https://pubchem.ncbi.nlm.nih.gov/compound/538757.
- 40 Rodin J O, Himel C M, silverstein R M, leeper R W and Gortner W A, Volatile flavor and aroma components of pineapple. I. Isolation and tentative identification of 2,5dimethyl-4-hydroxy-3(2H)-furanone, *J Food Sci*, 1965, **30**, 280–285.
- 41 Bijekar S R, Gayatri M C, Rajanna L, GC-MS profile of methanolic leaf extract of Baliospermum montanum (Wild.) Muell. Arg, *Int J Inno Res Sci Eng Technol*, 2015, 4(8).
- 42 Hiramoto K, Nasuhara A, Michikoshi K, Kato T and Kikugawa K, DNA strand-breaking activity and mutagenicity of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one (DDMP), a Maillard reaction product of glucose and glycine, *Mutat Res Genet Toxicol Environ Mutagen*, 1997, **395**(1), 47–56.
- 43 Kumar P P, Kumaravel S and Lalitha C, Screening of antioxidant activity, total phenolics and GC-MS study of Vitex negundo, *Afr J Biochem Res*, 2010, 4(7), 191-195.
- 44 Yu X, Zhao M, Liu F, Zeng S and Hu J, Identification of 2,3dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one as a strong antioxidant in glucose–histidine Maillard reaction products, Food Res Int, 2013, 51(1), 397-403.
- 45 Pubchem-2,3-Dihydrobenzofuran;https://pubchem.ncbi.nlm. nih.gov/compound/2_3-dihydrobenzofuran.
- 46 Gopalakrishnan S and Vadivel E, GC-MS analysis of some bioactive constituents of Mussaenda frondosa Linn., *Int J Pharma Bio Sci*, 2014, 2(1), 312-320.
- 47 Pubchem-2-Furancarboxaldehyde,5-(hydroxymethyl), https:// pubchem.ncbi.nlm.nih.gov/compound/5-hydroxymethylfurfural.
- 48 Xu C, Ou J, Cui Y, Wang L, Lv C et al., Development of a monoclonal antibody-based enzyme-linked immunosorbent assay for Tetrabromobisphenol A. *Monoclon Antib* Immunodiagn Immunother, 2013, **32**(2), 113-118.
- 49 Pubchem-Benzaldehyde,4-hydroxy-:https://pubchem.ncbi. nlm.nih.gov/compound/4-hydroxybenzaldehyde.
- 50 Pubchem- 2-Methoxy-4-vinylphenol: https://pubchem.ncbi.n lm.nih.gov/compound/2-Methoxy-4-vinylphenol.
- 51 Jeong J B, Hong S C, Jeong H J and Koo J S, Antiinflammatory effect of 2-methoxy-4-vinylphenol via the suppression of NF-κB and MAPK activation, and acetylation of histone H3, *Arch Pharm Res*, 2011, **34**, 2109.
- 52 Silici S and Kutluca S, Chemical composition and antibacterial activity of propolis collected by three different races of honeybees in the same region, *J Ethnopharmacol*, 2005, **99**(1), 69-73.
- 53 Pubchem-Phenol,2-methoxy-4-(2-propenyl); https://pubchem.ncbi.nlm.nih.gov/compound/eugenol
- 54 Kabuto H, Tada M and Kohno M, Eugenol [2-Methoxy-4-(2propenyl)phenol] prevents 6-hydroxydopamine- induced dopamine depression and lipid peroxidation inductivity in mouse striatum, *Biol Pharm Bull*, 2007, **30**(3), 423-427.
- 55 Chen Y, Zhou C, Ge Z, Liu Y, Liu Y et al., Composition and potential anticancer activities of essential oils obtained from myrrh and frankincense, *Oncol Lett*, 2013, 6(4), 1140-1146.
- 56 Hussain N, Abbasi T and Abbasi S A, Transformation of the pernicious and toxic weed parthenium into an organic fertilizer by vermicomposting, *Int J Environ Studies*, 2016, 73(5), 731-745.

- 57 Leal W S, Nakano Y, Kuwahara Y, Nakao H and Suzuki T, Pheromone study of acarid mites XVII: Identification of 2-Hydroxy-6-methyl benzaldehyde as the alarm pheromone of the Acarid Mite Tyrophagus perniciosus (Acarina: Acaridae), and its distribution among related mites, *Appl Entomol Zool*, 1988, **23**(4), 422-427.
- 58 Kuwahara Y, Sato M, Koshii T and Suzuki T, Chemical ecology of astigmatid Mites XXXII.2-Hydroxy-6-methylbenzaldehyde, the sex pheromone of the brown-legged grain mite Aleuroglyphus ovatus (TROUPEAU) (Acarina:acaridae), *Appl Entomol Zool*, 1992, 27(2), 253-260.
- 59 Pubchem-2-Hexadecen-1-ol,3,7,11,15-tetramethyl-,[R-[R*,R*-(E)]]- https://pubchem.ncbi.nlm.nih.gov/compound /5366244.
- 60 Olofsson P, Holmberg J, Tordsson J, Lu S, Åkerström B and Holmdahl R, Positional identification of Ncf1 as a gene that regulates arthritis severity in rats, *Nature genet*, 2003, 33(1), 25-32.
- 61 Pubchem-Hexadecanoicacid, methyl ester-https://pub chem.ncbi.nlm.nih.gov/compound/methyl_palmitate.
- 62 Pubchem-Hexadecanoic acid-https://pubchem.ncbi. nlm. nih.gov/compound/palmitic acid.
- 63 Yff B T S, Lindseya K L, Taylor M B, Erasmus D G and Jäger A K, The pharmacological screening of Pentanisia

prunelloides and the isolation of the antibacterial compound palmitic acid, *J Ethnopharmacol*, 2002, **79**(1), 101-107.

- 64 Gnanavel V and Sara A M, GC-MS analysis of petroleum ether and ethanol leaf extracts from Abrus precatorius LINN, *Int J Pharm Bio Sci*, 2013, **4**(3), 37-44.
- 65 Jananie R K, Priya V and Vijayalakshmi K, Determination of bioactive components of Cynodon dactylon by GC-MS analysis, *New York Sci J*, 2011, 4(4), 16-20.
- 66 Sudha T, Chidambarampillai S, Mohan V R, GC-MS analysis of bioactive components of aerial parts of Fluggea leucopyrus Willd. (Euphorbiaceae), *J Appl Pharm Sci* 2013, 3(05), 126-130.
- 67 Mujeeb F, Bajpai P, Pathak N, Phytochemical evaluation, antimicrobial activity and determination of bioactive components from leaves of Aegle marmelos, *BioMed Res Int*, 2014, 1.
- 68 Osawa, T, Novel natural antioxidants for utilization in food and biological systems. In Posthar Biochem plant Food-Mater Trop; Uritani, I, Garcia V V, Mendoza E M, eds, (Japan Scientific Societies Press, Tokyo, Japan), 1994, 241-251.
- 69 Zheng W and Wang S Y, Antioxidant activity and phenolic compounds in selected herbs, *J Agric Food Chem*, 2001, 49, 5165–5170.